

S15.12 Inhibition of mammalian cytochrome *bc*₁ complex by chromanols and related compounds

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Tocopherols (chromanols) and tocopheryl quinones occur naturally in biological membranes. Related 6-hydroxy-chromanones were observed as byproducts of industrial tocopherol processing. Because of the extensive use of some tocopherols as food supplements, the aim of the study was to assess their inhibiting properties at the mammalian cytochrome *bc*₁ complex as toxicological and pharmacological target. The effects of these compounds and low molecular analogues at the isolated cytochrome *bc*₁ complex from bovine heart were studied by dual-wavelength photometry, stopped flow photometry, and low temperature EPR spectroscopy. Our data show that α - and γ -tocopherols as well as α -tocopheryl quinone required millimolar concentrations to achieve half inhibition (IC₅₀) of the decylubiquinol: cytochrome *c* oxidoreductase activity. In contrast, γ -tocopheryl quinone and the less lipophilic 6-hydroxy-4,4,7,8-tetramethyl-chroman-2-one (TMC2O) exhibited IC₅₀ values of 117 ± 6 and 94 ± 12 μ M, respectively. Stopped flow measurements of the reduction of cytochromes in the isolated cytochrome *bc*₁ complex revealed that TMC2O preferably inhibited the cytochrome *c*₁ reduction. EPR spectra of the ascorbate-reduced Rieske iron sulfur protein at 20 K documented a shift of g_z from 2.0288 ± 0.0005 (control) to 2.02240 ± 0.0003 (TMC2O) indicating a specific binding of this compound. Besides toxicological implications for the effect of γ -tocopheryl quinone on the cytochrome *bc*₁ activity in vivo, our data suggest that synthetic modifications of TMC2O could lead to a new class of cytochrome *bc*₁ inhibitors.

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S15.13 Is cytochrome *b* glutamic acid 272 a quinol binding residue in the *bc*₁ complex?

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On the basis of the three-dimensional structures of the *bc*₁ complex in the presence of the inhibitor stigmatellin, it is assumed that the substrate quinol binding involves the cyt *b* glutamate residue E272. However, this residue is not totally conserved during evolution: in some β and γ -proteobacteria, it is replaced by valine or proline and another glutamate is conserved and corresponds to H253 in yeast. Studies of *bc*₁ complexes with E272/H253 mutations will be presented. Replacement of E272 with apolar residues did not abolish the *bc*₁ activity, although it slowed down the kinetics of electron transfer. The *K*_m value for the binding of the substrate quinol was not modified, and the EPR data showed that the quinol binding still occurred in the mutants. Binding of stigmatellin was retained, however mutations E272P/V induced resistance toward myxothiazol. The pH dependence of the *bc*₁ activity was not modified in the absence of the glutamate E272. Our results suggest that residue E272 may not be involved in direct substrate binding or in its direct deprotonation. Revertants were selected with polar residues serine and threonine at position 272. The data lead us to suggest that E272 may be involved in a later

step on the proton exit pathway via the interaction with a water molecule.

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S15.14 Stochastic approach of *bc*₁ complex functioning

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The *bc*₁ complex is a central complex in the mitochondrial respiratory chain. It links the transfer of electrons from ubiquinol (QH₂) to cytochrome *c* and proton translocation across the inner mitochondrial membrane. It is widely agreed that the “Q-cycle mechanism” proposed by Mitchell correctly describes the *bc*₁ complex functioning. It is based on an unexpected separation of the two electrons coming from the QH₂ molecule bound at the Q_o site of the *bc*₁ complex. One electron is transferred to the iron–sulfur centre (FeS) of the iron sulphur protein (ISP) and the second to the lower potential heme b_L. The electron on heme b_L moves within the cytochrome *b* to reduce the higher potential heme b_H, which in turn reduces an ubiquinone (Q) or a semiquinone (SQ) at a second ubiquinone binding site Q_i. Using a stochastic approach based on the known spatial structure of *bc*₁ complexes and the kinetic parameters described by Moser and Dutton, we have demonstrated the natural emergence of the Q-cycle mechanism and the quasi absence of short-circuits in the functional dimer of *bc*₁ complex without the necessity to invoke any additional mechanism. In this poster we studied the various parameters which influence the reaction between redox centers. The natural distances measured on the crystallographic structures appeared to minimize the short-circuits thus favouring an energetic “Q-cycle” mechanism.

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S15.15 Remodelling *bc*₁ complex Q_o site in yeast to study acquired resistance to inhibitors

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Inhibitors of the mitochondrial respiratory chain enzyme cytochrome *bc*₁ – especially inhibitors targeting the Q_o site of the enzyme – have been developed as anti-microbial agents. They are used in agriculture to control plant pathogenic fungi and in medicine against human pathogens, such as the malaria parasite. Unfortunately, the problem of acquired resistance has rapidly emerged. Resistance is often linked to mutation within the mitochondrially-encoded cytochrome *b* that forms the Q_o site. To obtain information on the molecular basis of the resistance and of the differential evolution of the mutations, we used yeast (*Saccharomyces cerevisiae*) as a model organism. We modified the Q_o site of yeast *bc*₁ complex and introduced variations and resistance mutations found in pathogens. The respiratory activity, the sensitivity to inhibitors and the fitness of the yeast mutants were then analyzed. In particular, we studied the resistance mutations F129L, G137R and G143A found in many pathogenic fungi of important crops. We addressed the question of the impact of structural variations in the Q_o site and of variations in the intron/exon structure of the cytochrome *b* gene on the evolution of G143A. We also remodeled